

# Raster Image Correlation Spectroscopy RICS

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# Raster Image Correlation Spectroscopy

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We can have a combination of very high time resolution with sufficient spatial resolution.

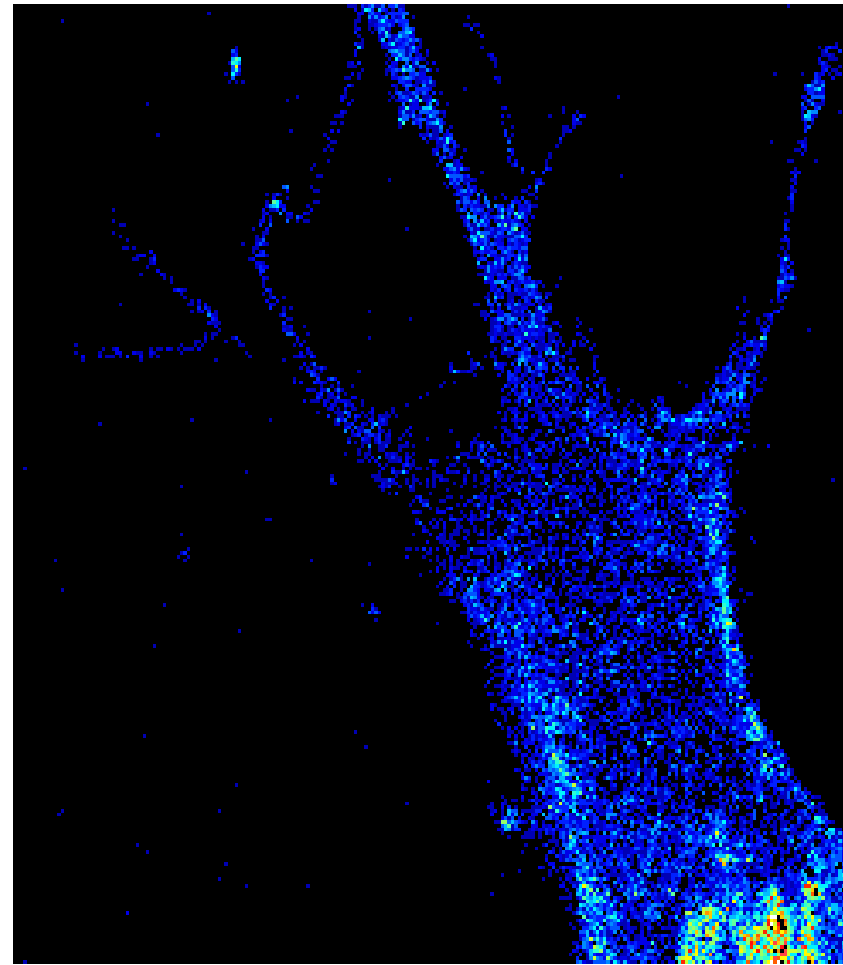
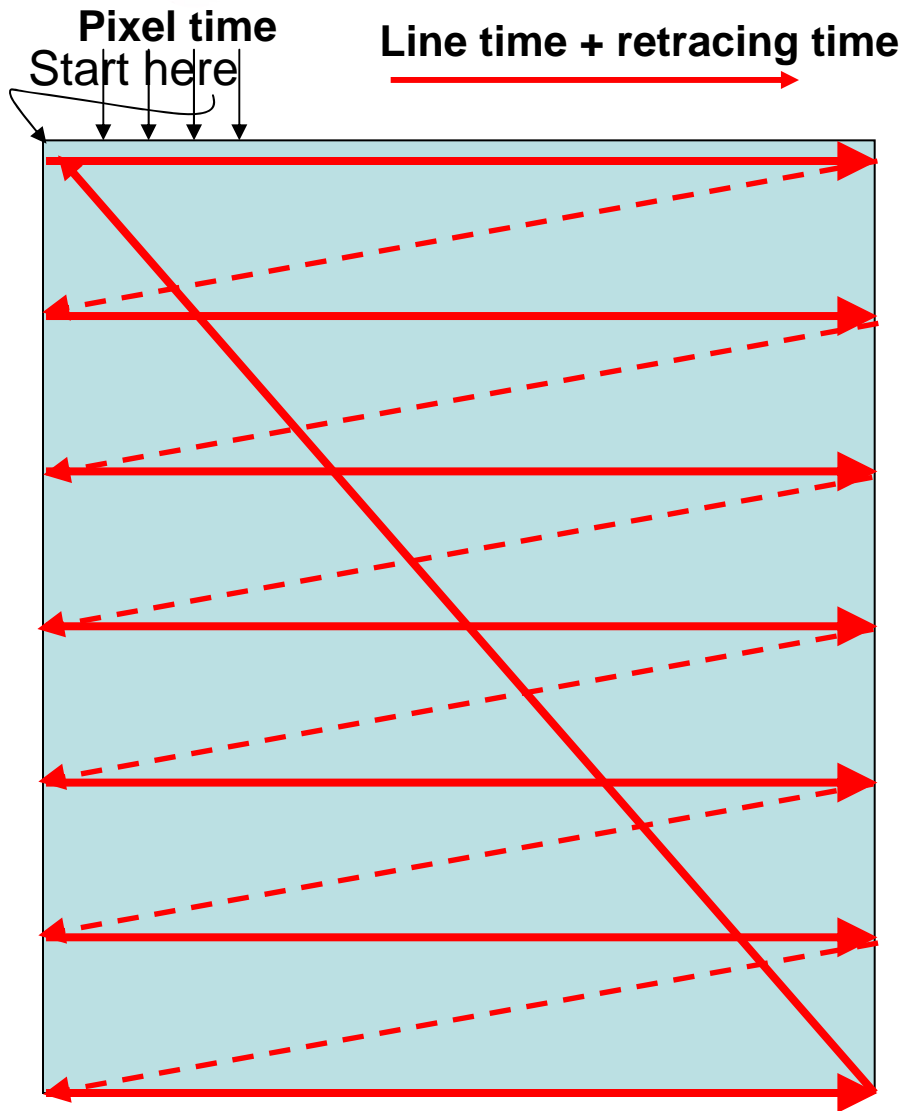
## Major benefits of RICS:

- It can be done with **commercial laser scanning microscopes** (either one or two photon systems)
- It can be done with **analog detection**, as well as with photon counting systems, although the characteristic of the detector must be accounted for (time correlations at very short times due to the analog filter)
- RICS provides an intrinsic method to separate the immobile fraction
- It provides a powerful method to distinguish diffusion from binding

How does it work?

# Raster Scanning

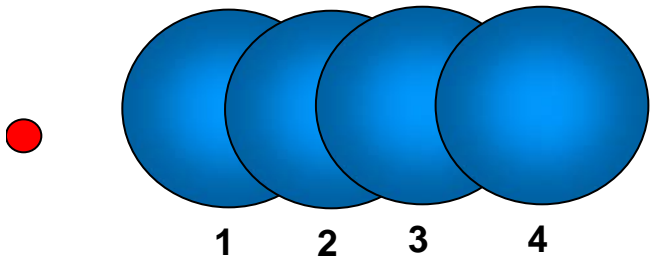
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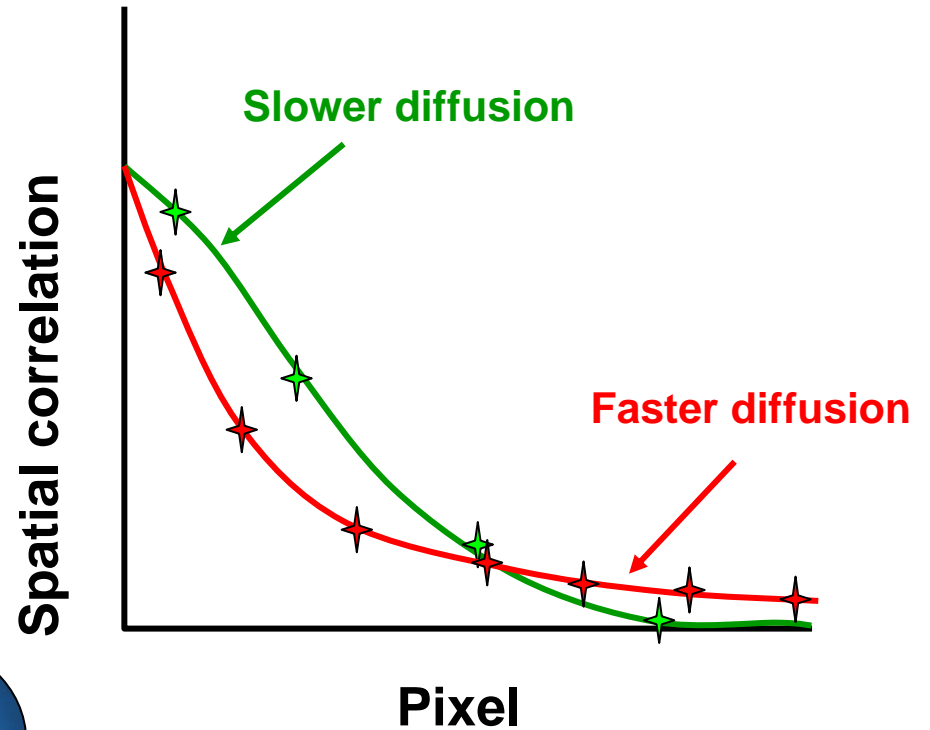
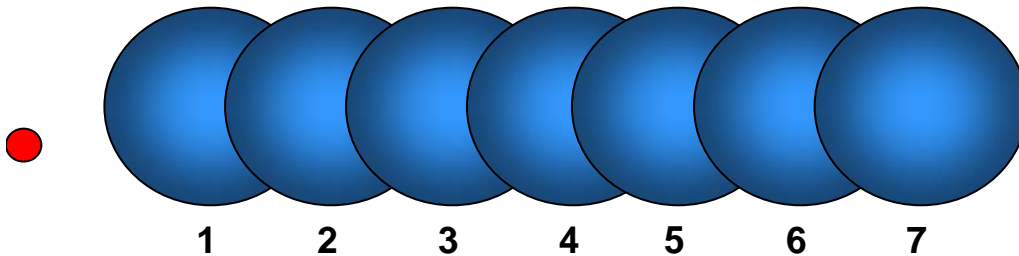
# Temporal information hidden in the raster-scan image: the RICS approach

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Situation 1: slow diffusion

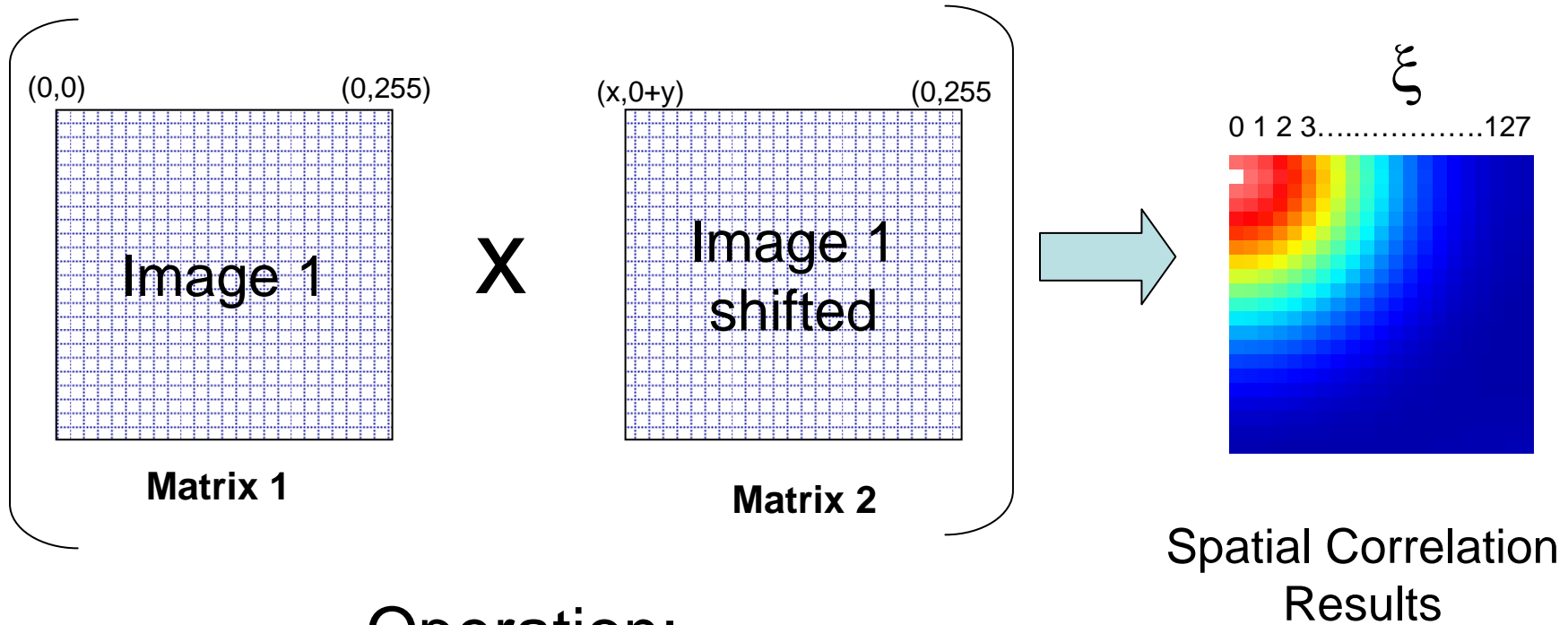


Situation 2: fast diffusion



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# How is the spatial correlation done?



## Operation:

In the x direction

PLUS In the y direction

$$(0,0 \times 0,0) + (0,1 \times 0,1) + (0,2 \times 0,2) \dots (0,127 \times 0,127) \\ + (1,0 \times 1,0) + (1,1 \times 1,1) + (1,2 \times 1,2) \dots (1,127 \times 1,127)$$

One number is obtained for x and y and is divided by the average intensity squared



# The RICS approach: 2-D spatial correlations

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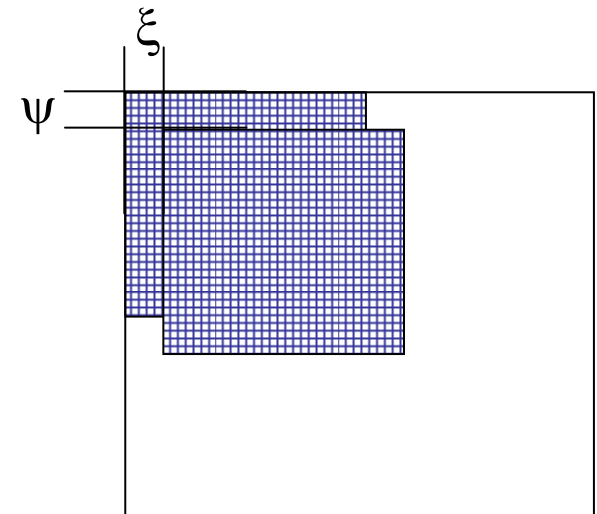
In a raster-scan image, points are measured at different positions and at different times simultaneously

If we consider the **time sequence**, it is not continuous in time

If we consider the **pixel sequence**, it is contiguous in space

In the RICS approach we calculate the 2-D spatial correlation function (similarly to the ICS method of Petersen and Wiseman)

$$G_{RICS}(\xi, \psi) = \frac{\langle I(x, y)I(x + \xi, y + \psi) \rangle}{\langle I(x, y) \rangle^2} - 1$$



The variables  $\xi$  and  $\psi$  represent spatial increments in the x and y directions, respectively

2-D spatial correlation can be computed very efficiently using FFT methods.

To introduce the “RICS concept” we must account for the relationship between time and position of the scanning laser beam.

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# The RICS approach for diffusion

The dynamic at a point is independent on the scanning motion of the laser beam

$$G_{RICS}(\xi, \psi) = S(\xi, \psi) \times G(\xi, \psi)$$

Consider now the process of diffusion. The diffusion kernel can be described by the following expression

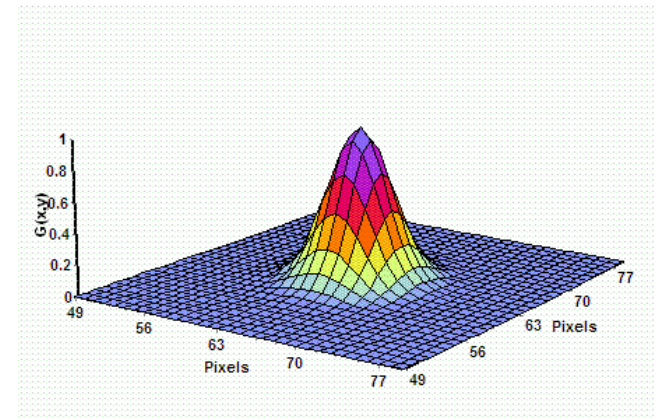
$$P(r, t) = \frac{1}{(4\pi Dt)^{3/2}} \exp\left(-\frac{r^2}{4Dt}\right)$$

There are two parts:

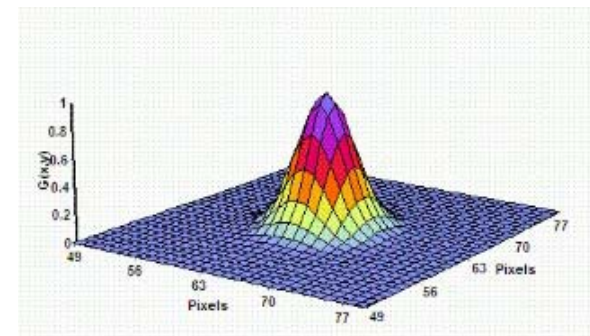
- (1) the temporal term,
- (2) the spatial Gaussian term

For any diffusion value the amplitude decreases as a function of time and the width of the Gaussian increases as a function of time

**FAST**



**SLOW**



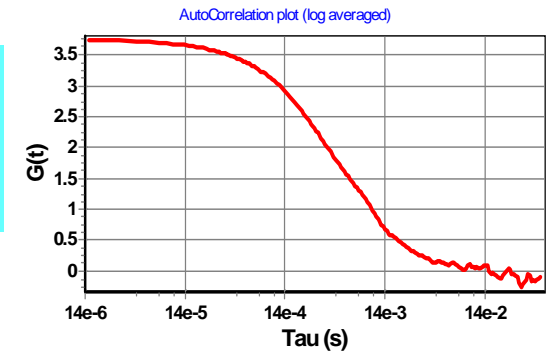




# RICS: space and time relationships

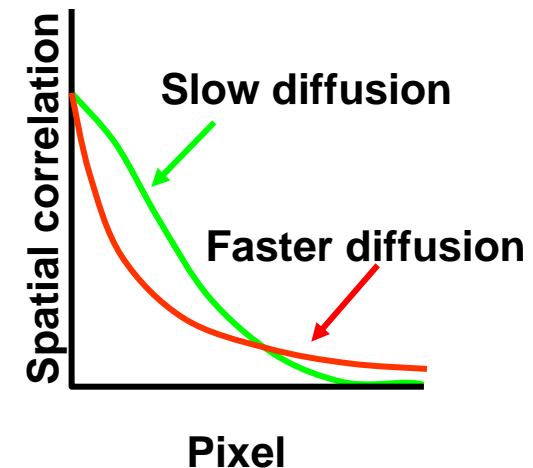
At any position, the ACF due to diffusion takes the familiar form:

$$G(\xi, \psi) = \frac{\gamma}{N} \left( 1 + \frac{4D(\tau_p \xi + \tau_l \psi)}{w_0^2} \right)^{-1} \left( 1 + \frac{4D(\tau_p \xi + \tau_l \psi)}{w_z^2} \right)^{-1/2}$$



$\tau_p$  and  $\tau_l$  indicate the pixel time and the line time.  
The correlation due to the scanner movement is:

$$S(\xi, \psi) = \exp \left( - \frac{\left[ \left( \frac{2\xi \delta r}{w_0} \right)^2 + \left( \frac{2\psi \delta r}{w_0} \right)^2 \right]}{\left( 1 + \frac{4D(\tau_p \xi + \tau_l \psi)}{w_0^2} \right)} \right)$$



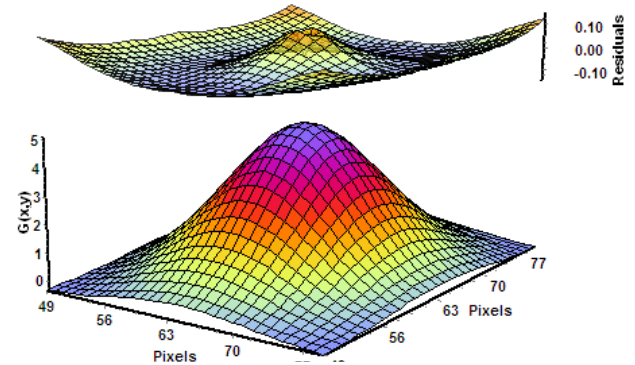
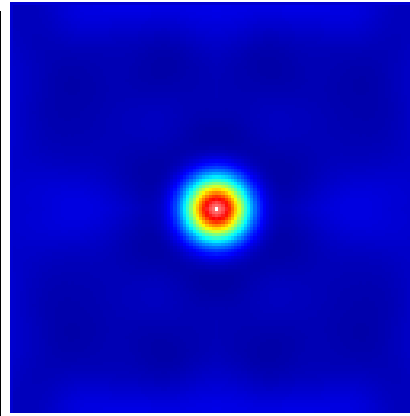
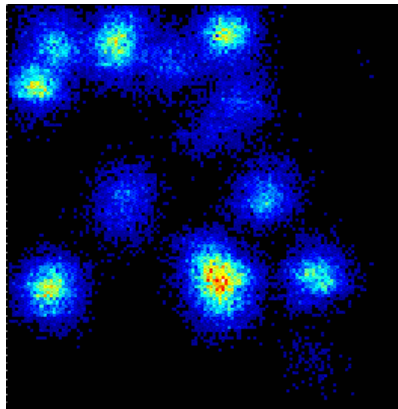
Where  $\delta r$  is the pixel size. For  $D=0$  the spatial correlation gives the autocorrelation of the PSF, with an amplitude equal to  $\gamma/N$ . As  $D$  increases, the correlation ( $G$  term) becomes narrower and the width of the  $S$  term increases.

# RICS Simulations of three different diffusion rates:

Box size=3.4 $\mu\text{m}$  sampling time: 1) 32 $\mu\text{s}$ /pixel 2) 8 $\mu\text{s}$ /pixel 3) 4 $\mu\text{s}$ /pixel

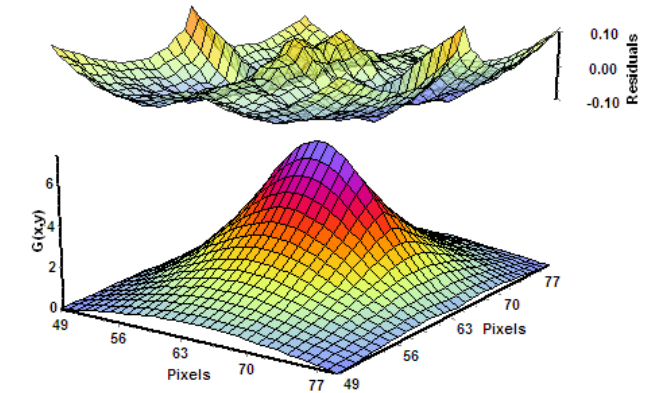
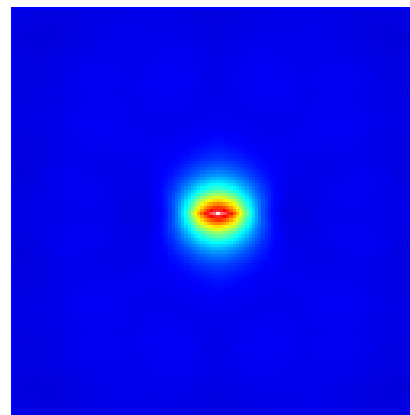
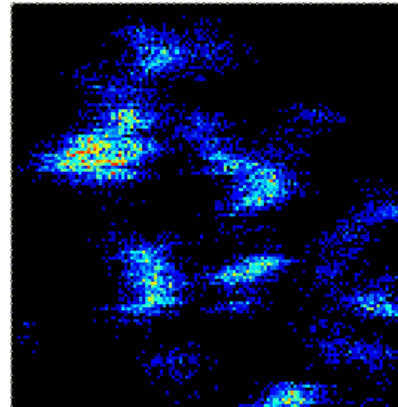
**$D = 0.1 \mu\text{m}^2/\text{s}$**

(membrane proteins)



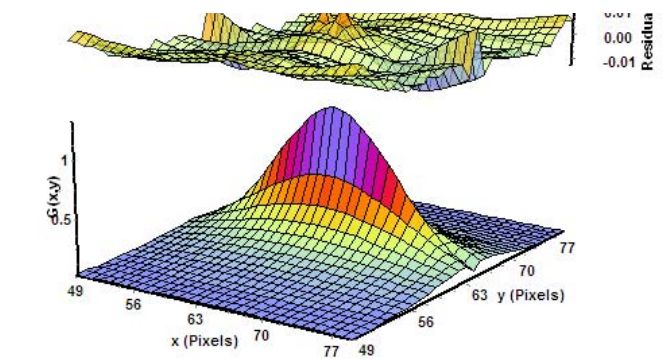
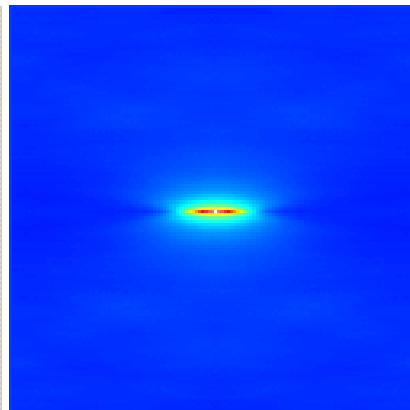
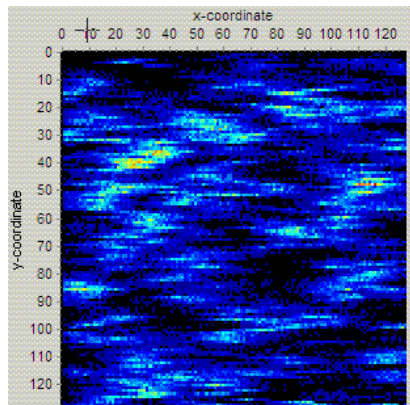
**$D = 5.0 \mu\text{m}^2/\text{s}$**

(40 nm beads)



**$D = 90 \mu\text{m}^2/\text{s}$**

(EGFP)

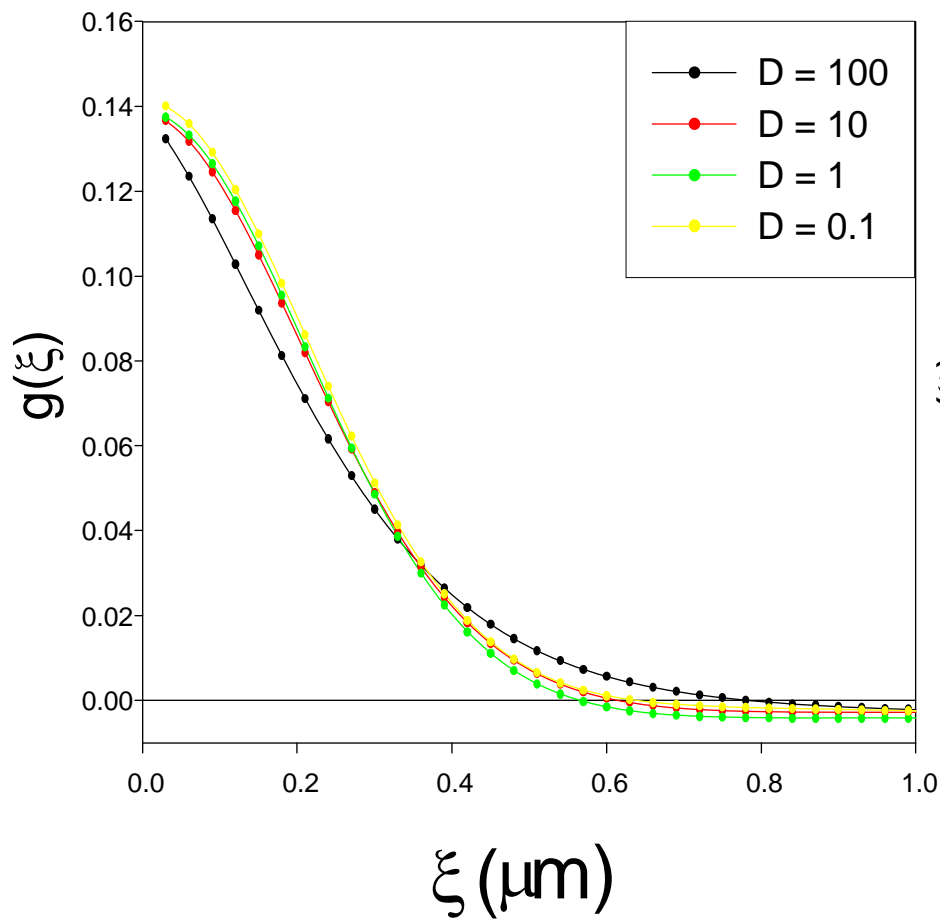


lf<sub>d</sub>

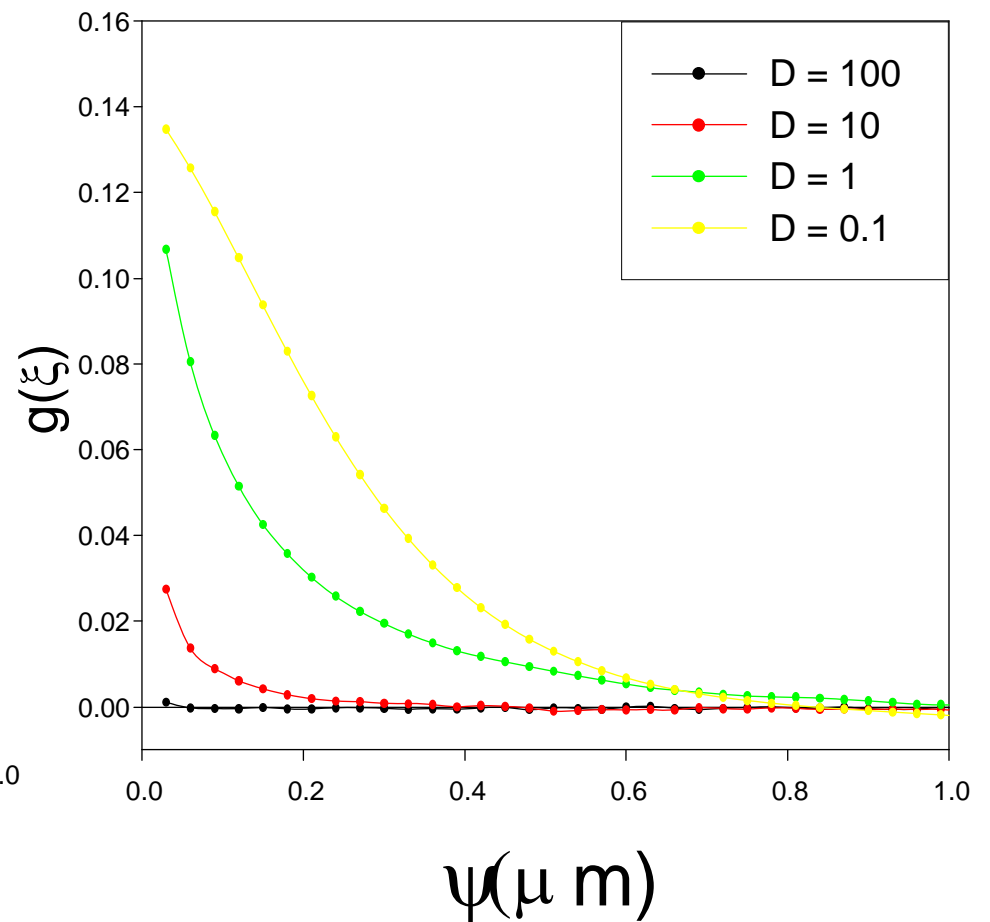
# Horizontal and Vertical fits:

Simulations of beads 300 frames, 128x128pixels, 8μs/pix, size of pixels=30nm

## Horizontal ACF



## Vertical ACF



In SIMFCS

# How to Setup the Laser Scanning Confocal Microscope

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## ● Scan Speeds ( $\mu\text{s}/\text{pixel}$ ):

- $4\mu\text{s}$  for fast molecules  $D > 100\mu\text{m}^2/\text{s}$
- $8 - 32\mu\text{s}$  for slower molecules  $D = 1\mu\text{m}^2/\text{s} - 100\mu\text{m}^2/\text{s}$
- $32 - 100\mu\text{s}$  for slower molecules  $D = 0.1\mu\text{m}^2/\text{s} - 10\mu\text{m}^2/\text{s}$

## ● Pixel Size:

- 3-4x smaller than the Point Spread Function (PSF  $\sim 300\text{nm}$ )

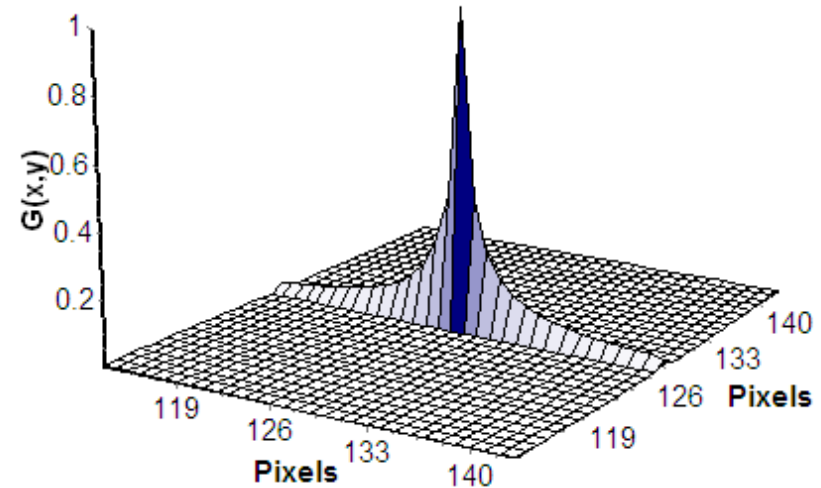
## ● Molecular Concentrations

- Same conditions as conventional FCS methods

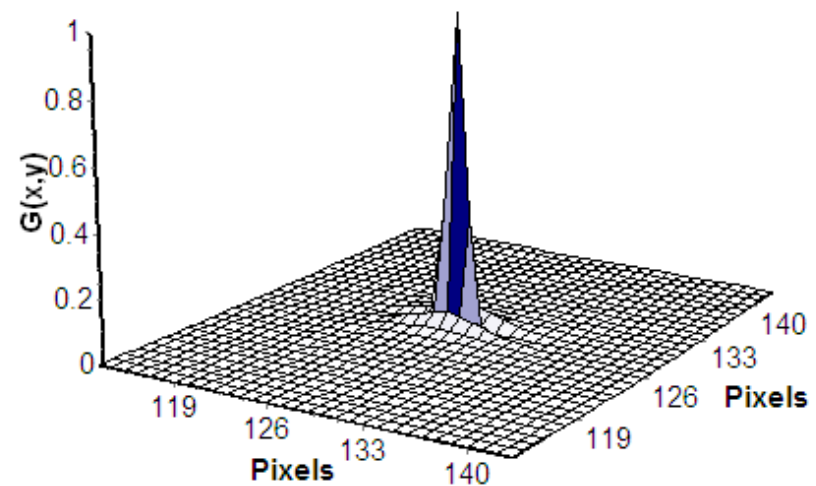
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# Common Errors in RICS

**Scanning Too Slow**  
(100  $\mu\text{s}/\text{pixel}$ ,  $D = 300 \mu\text{m}^2/\text{s}$ )



**Pixels are separated too much**  
relative to PSF  
(pixel size =  $w_0 = 0.3 \mu\text{m}$ )

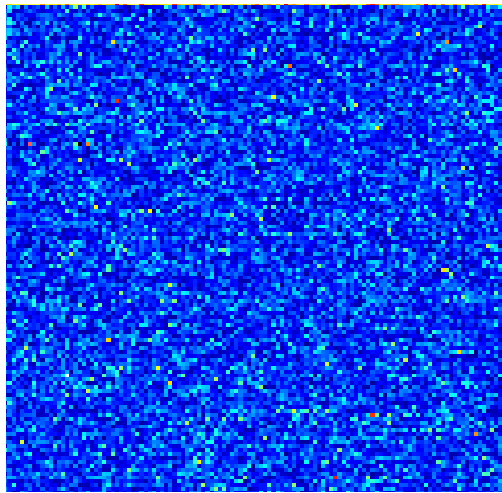


Courtesy of Jay Unruh

# RICS: Fits to spatial correlation functions

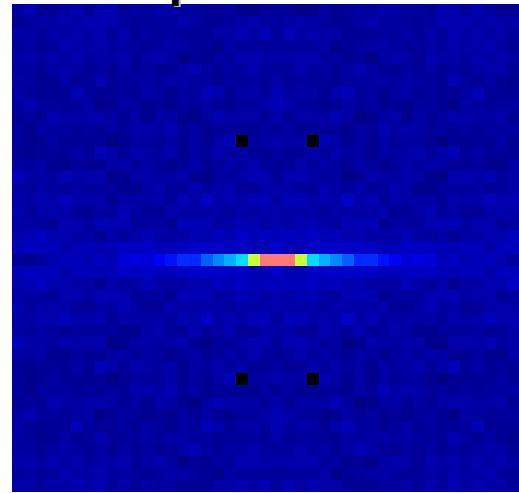
Olympus Fluoview300 LSM

EGFP in solution

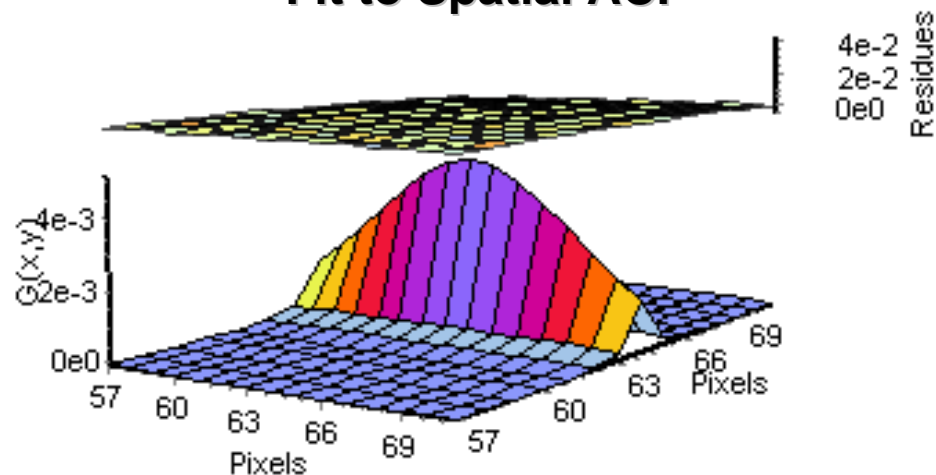


128x128, 4  $\mu\text{s}/\text{pixel}$ , 5.4 ms/line, 0.023  $\mu\text{m}/\text{pixel}$

Spatial ACF

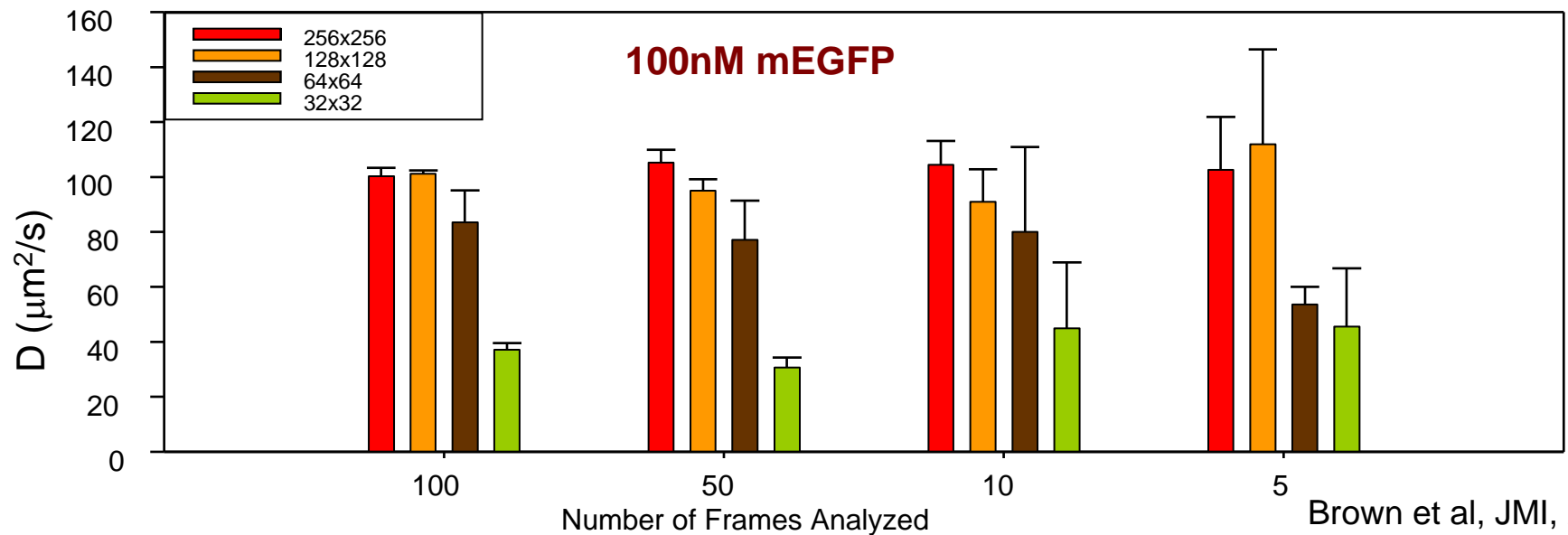
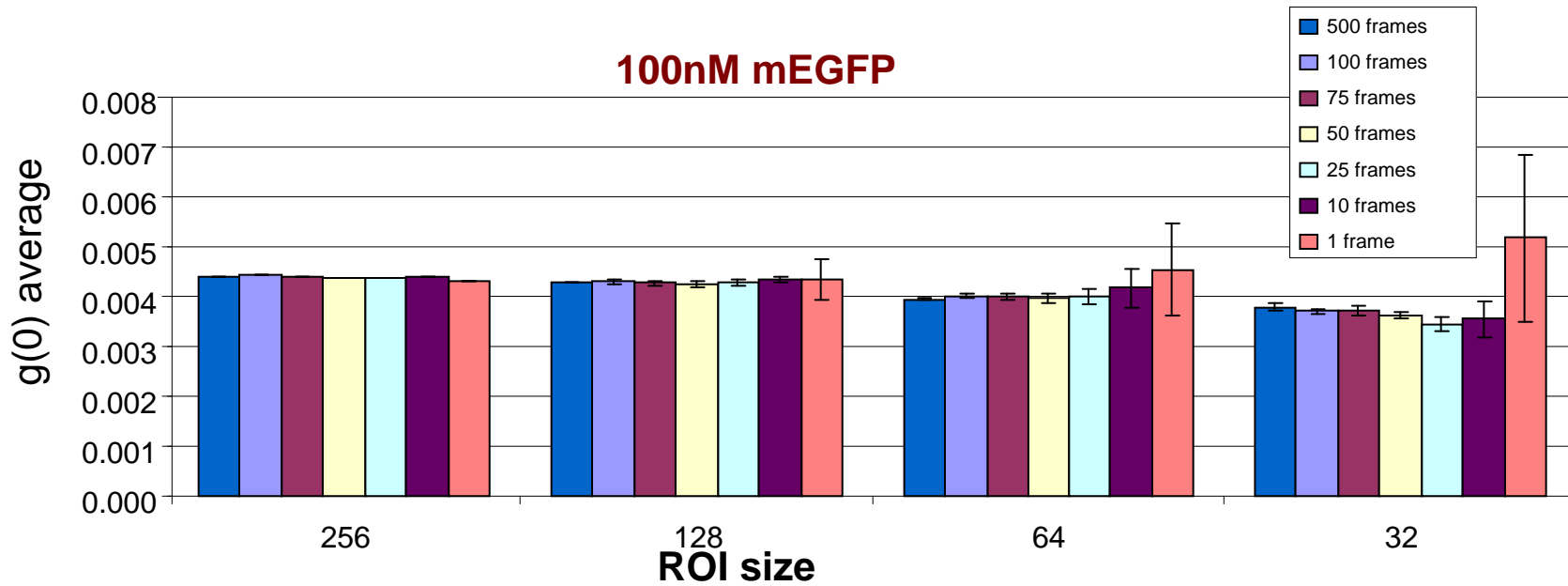


Fit to Spatial ACF

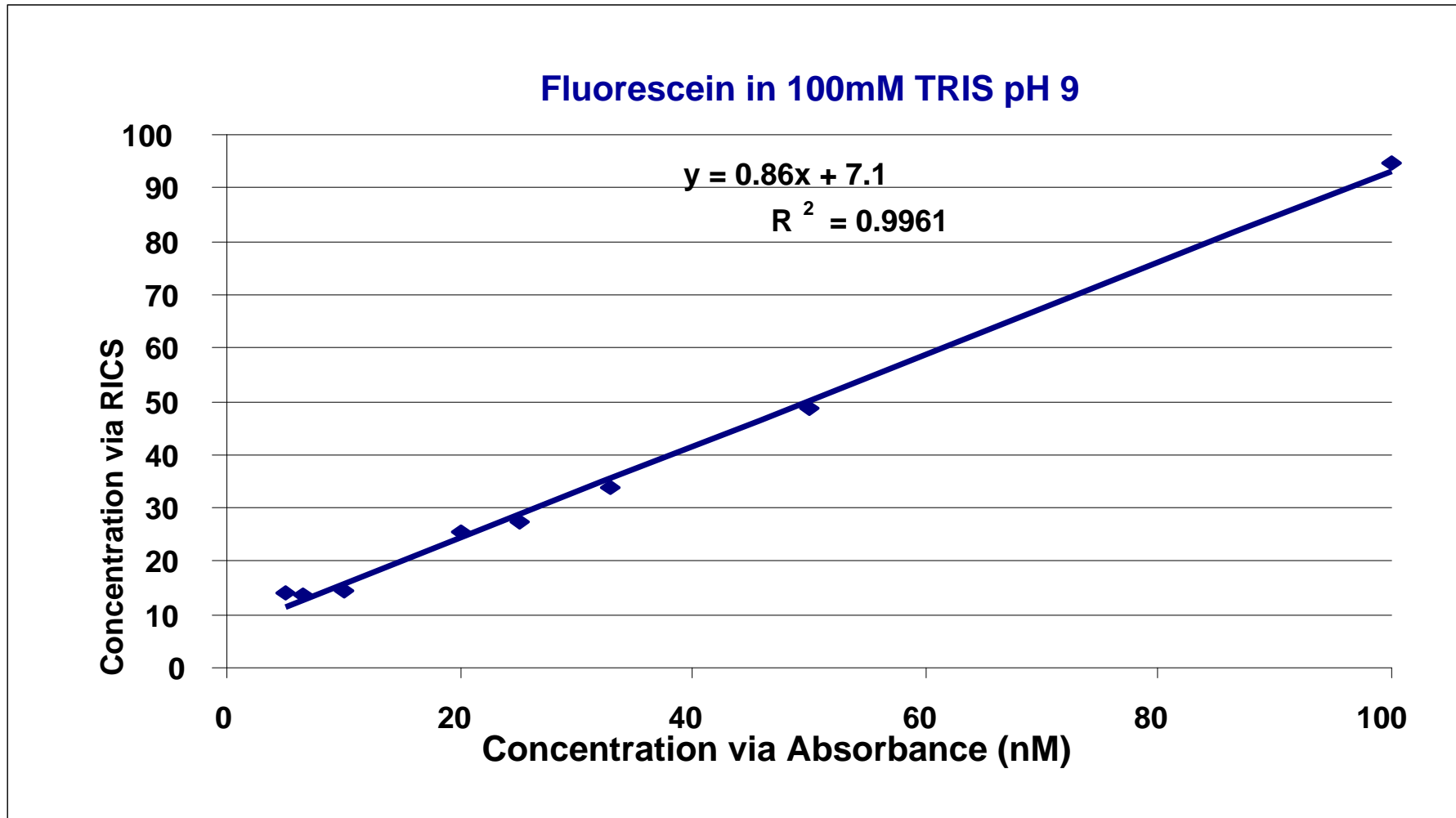


$$D = 105 \pm 10 \mu\text{m}^2/\text{s}$$

# What ROI size to use? How many frames to acquire?



# Obtaining concentration from RICS





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## How we go from solutions to cells?

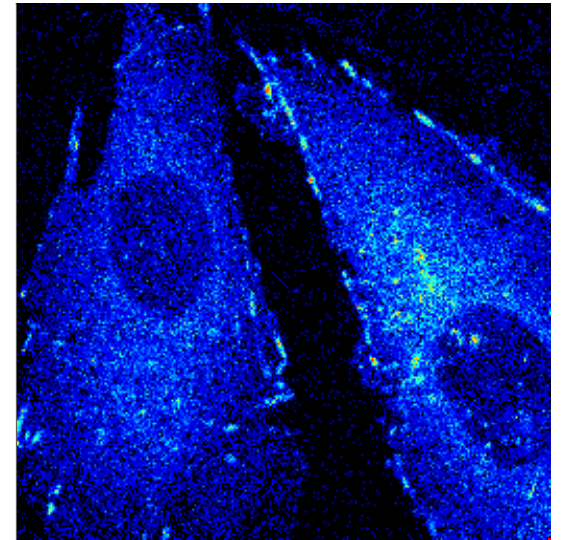
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In cells we have an **immobile fraction**

The 2-D-spatial correlation of an image containing immobile features has a very strong correlation pattern

We need to separate this **immobile** fraction from the mobile part before calculating the transform

How is this achieved?



# Does noise from the detectors correlate?

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In a “truly immobile” bright region, the intensity fluctuates according to the Poisson distribution due to shot noise.

The time correlation of the shot noise is zero, except at time zero.

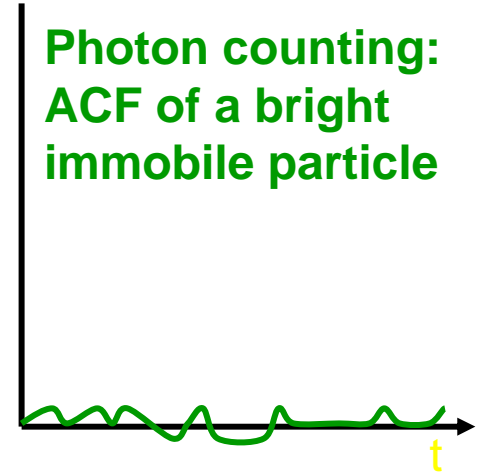
The spatial correlation of the intensity at any two pixels due to shot noise is zero, **even if the two points are within the PSF.**

If we subtract the average intensity and disregard the zero time-space point, the immobile bright region **totally disappear** from the correlation function

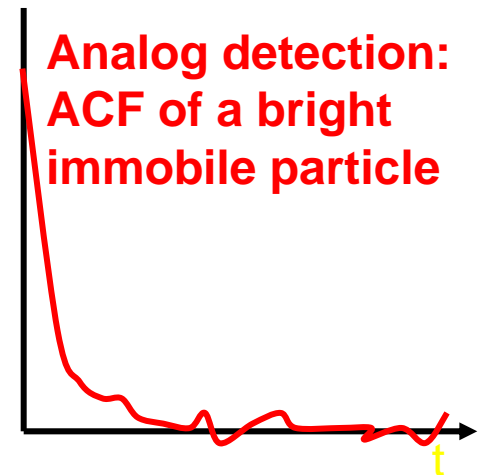
## Attention!!!!

This is not true for analog detection, not even in the first order approximation. For analog detection the shot noise is time (and space) correlated.

Photon counting:  
ACF of a bright  
immobile particle

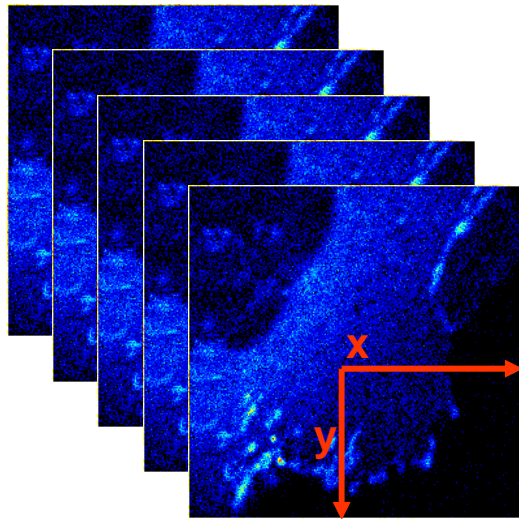


Analog detection:  
ACF of a bright  
immobile particle

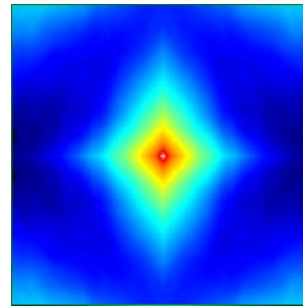


# Formula used to subtract background:

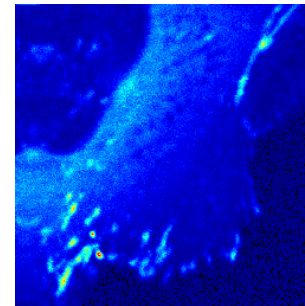
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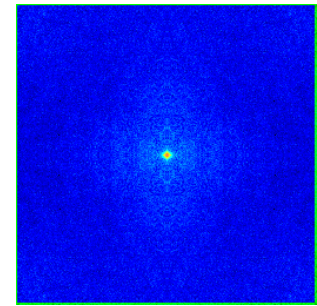
Spatial Correlation



Spatial correlation before subtracting background



Subtract the average



Spatial Correlation of entire image After subtracting image

Average intensity of each pixel on the overall stack:  $\overline{I(x, y)}$

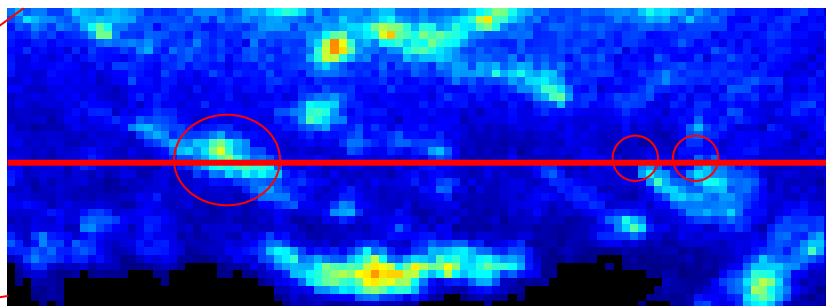
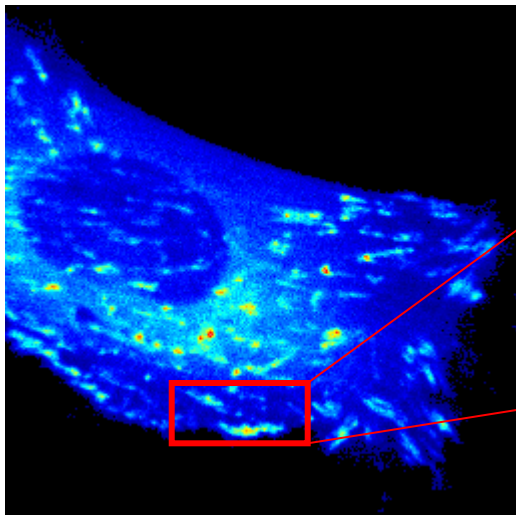
$I_i(x, y) - \overline{I(x, y)}$  The intensity of each pixel minus the average intensity from entire stack for each pixel: However, this yields negative values.

A scalar must be added :  $a = \overline{I}$

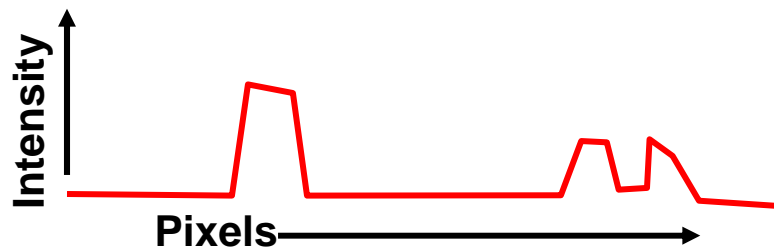
$$ICS(F_i(x, y)) \quad \text{where} \quad F_i(x, y) = I_i(x, y) - \overline{I(x, y)} + a$$

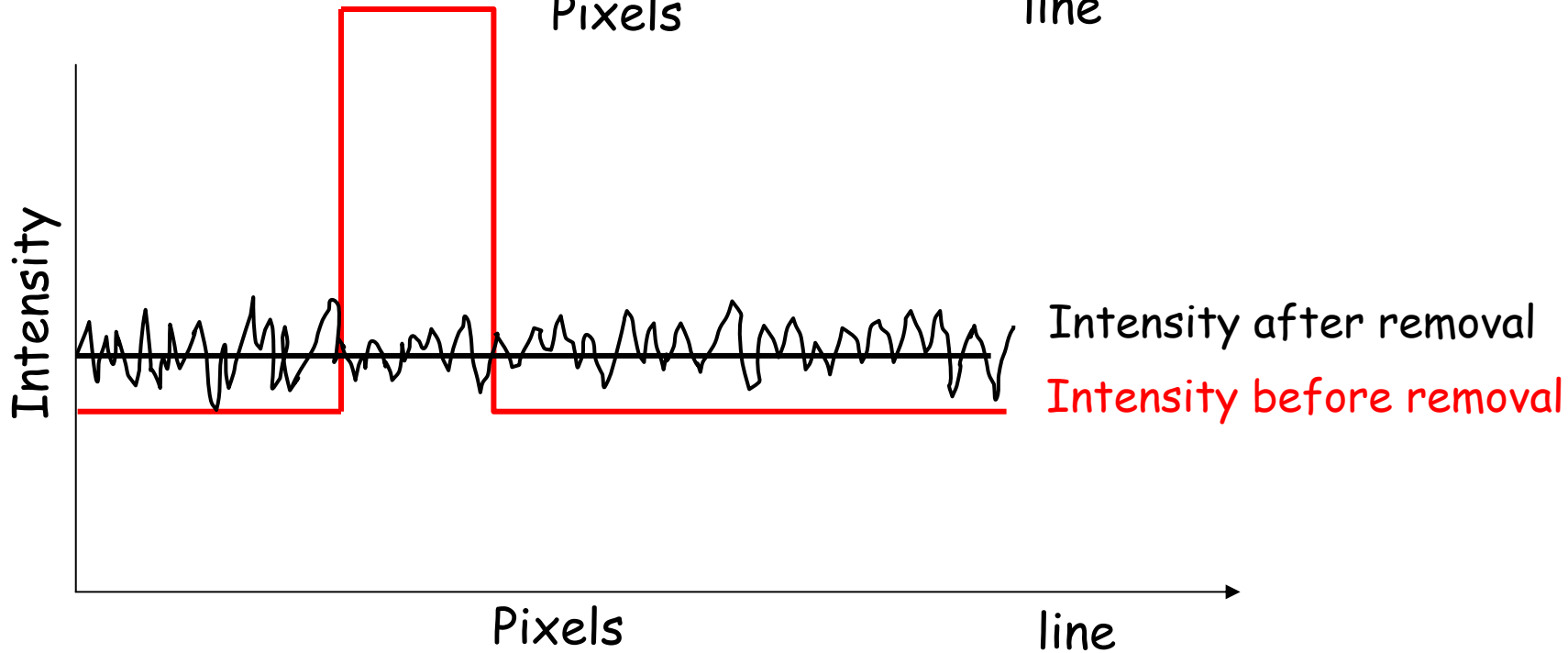
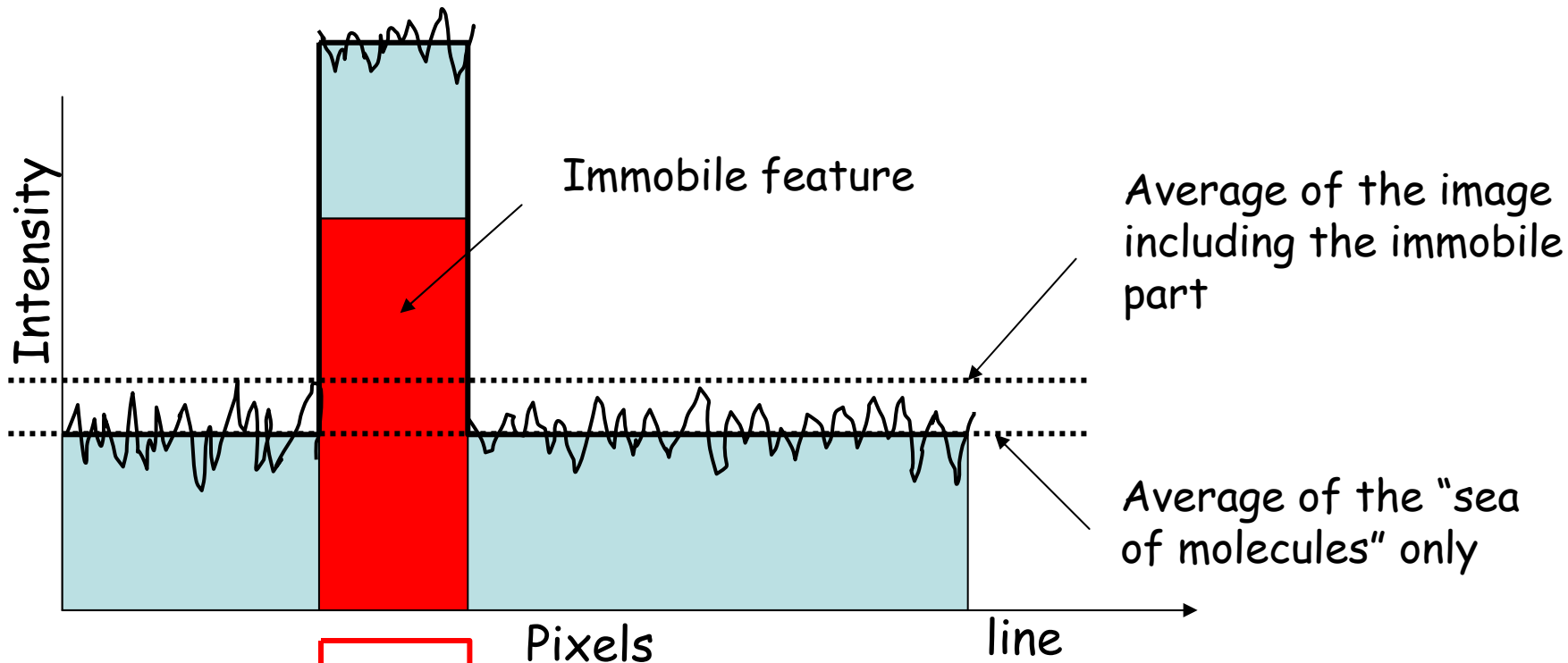
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# How to subtract immobile features from images?



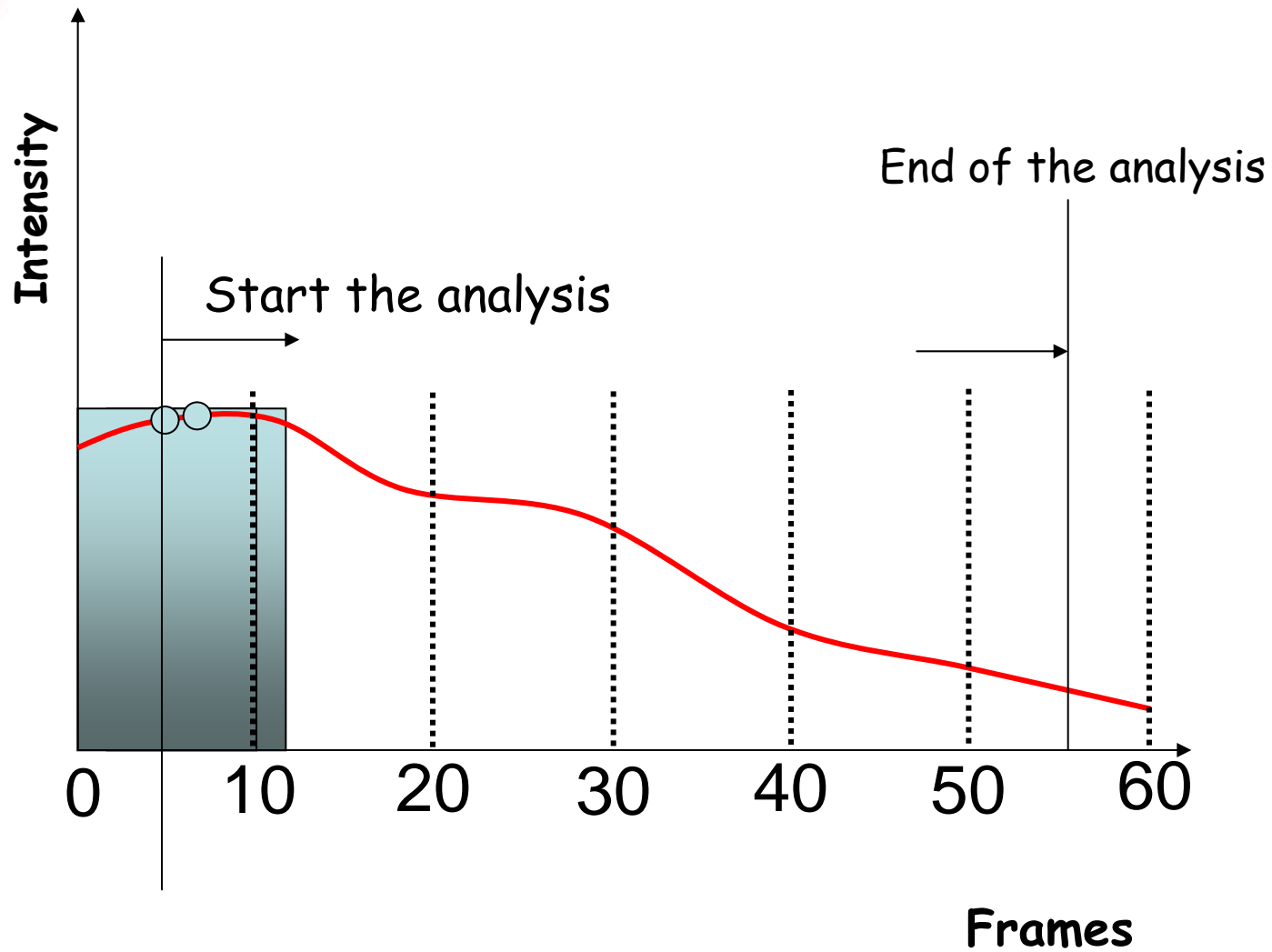
Intensity profile





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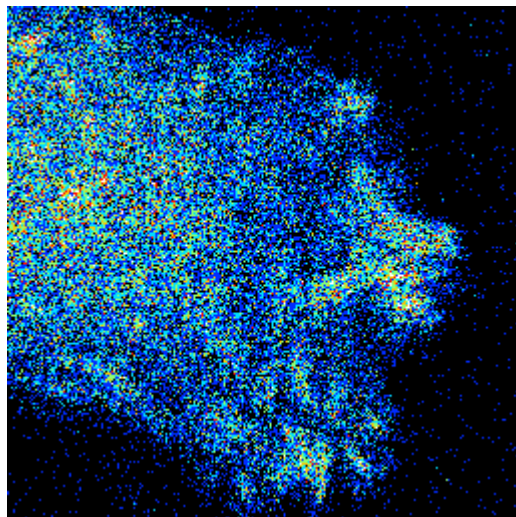
# Subtraction of moving average



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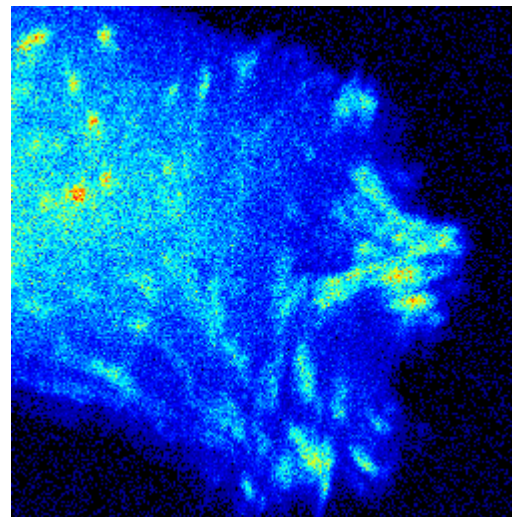
# Moving average operation on frames:

Frame #5

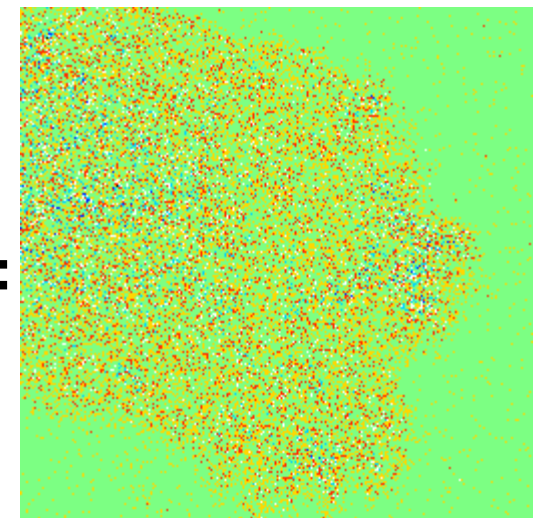


Matrix 1

Average  
between 1-10



Matrix 2



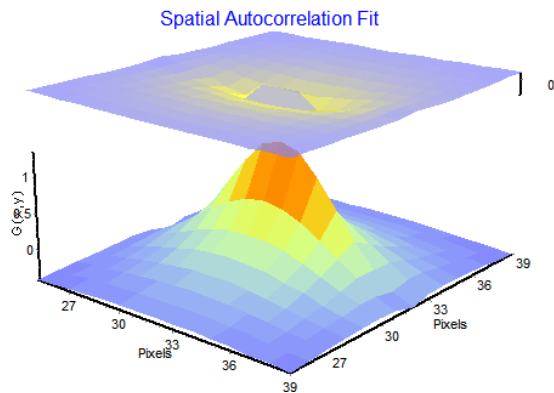
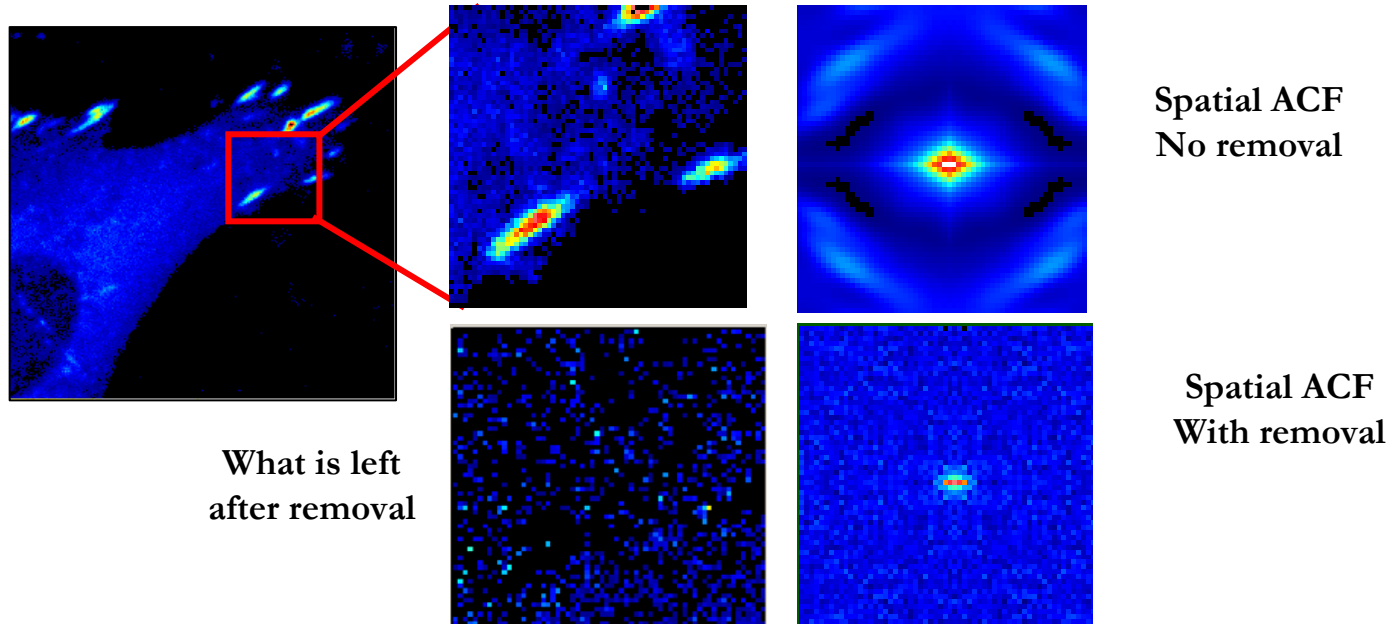
A scalar average is then added

Operation is repeated for frame #6 - average between 2-11  
frame #7 - average between 3-12



# Example of the Removal of Immobile Structures and Slow Moving Features

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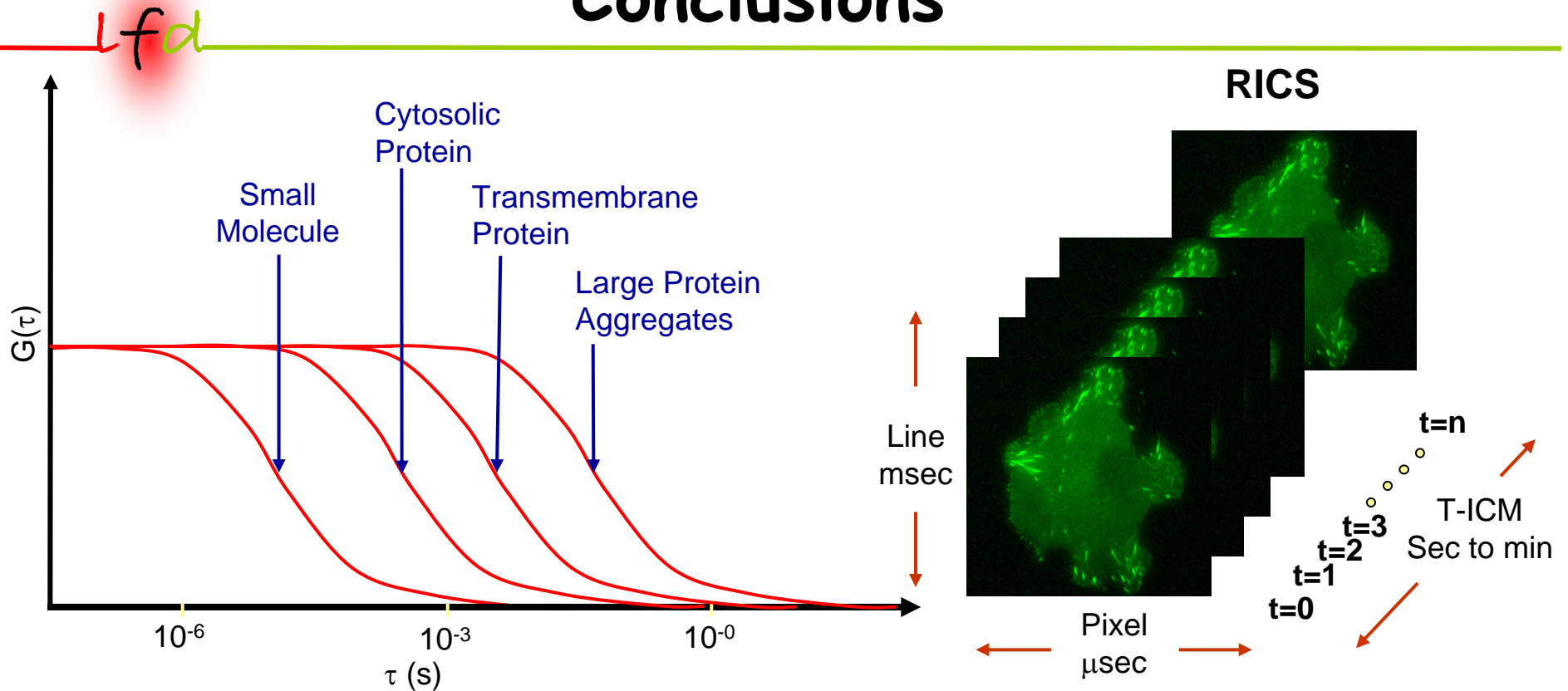
Fit using 3-D diffusion formula

Pixel size =  $0.092\mu\text{m}$   
 Pixel time =  $8\mu\text{s}$   
 Line time =  $3.152\text{ms}$   
 $W_0$  =  $0.35\mu\text{m}$

$G1(0)$	=	0.0062
D1	=	$7.4\mu\text{m}^2/\text{s}$
$G2(0)$	=	0.00023
D2	=	$0.54\mu\text{m}^2/\text{s}$
Bkgd	=	-0.00115



# Conclusions



Techniques	Time Res.	Spatial Res.	Used to Study
<b>FCS</b>	sec	<0.5 $\mu$ m	Protein aggregates Transmembrane proteins
Temporal ICM	sec	<0.5 $\mu$ m	Protein aggregates Transmembrane proteins
<b>RICS</b>	$\mu$ sec-msec	~2 $\mu$ m	Soluble proteins Binding interactions
RICS	$\mu$ sec-msec	~2 $\mu$ m	Soluble proteins Binding interactions
Line-RICS	msec	<0.5 $\mu$ m	Soluble proteins Binding interactions

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# Summary of RICS

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- Measures dynamic rates from the  $\mu\text{sec}$ -sec time scale
- Anyone with a commercially available instrument can use it
- Immobile structures can be filtered out and fast fluctuations can be detected
- RICS has high spatial and temporal resolution
- The range of these dynamic rates covers a wide range from immobile to cytosolic diffusions ( $0.001\text{-}300\mu\text{m}^2/\text{s}$ )
- Other types of processes and interactions are also measured
- Line scanning is essential for determination of binding process and complements the RICS analysis